Serological Evaluation Of The Presence Of Anti-Spermatozoa Antibody In Infertile Males In Thi-Qar Governorate

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Abstract

A case control study was conducted in Thi-Qar governorate, south of Iraq, included a total 154 individuals 104 of theme were infertile patients while the other 50 males were fertile healthy control males, who attended the infertility center in A Al-Hussein Teaching +0hospital, from 20/10/2016 to 16/1/2018. aged between (18-51) years with mean of age was (29.95 ± 0.68) and (50) healthy controls males aged between (23-37) years with mean age was (27.58±0.51). All patient submitted full reports about their medical status history, in addition to their infertility type (primary or secondary). Seminal fluid samples were collected by masturbation after three days of sexual avoidance. All patients and healthy control males were tested for presence of anti-spermatozoa antibodies in their seminal fluid. seminal fluid analysis in this study showed there were significant differences (p < 0.05) between fertile control group and infertile males (patients) in (The seminal fluid viscosity, Liquefaction time, Semen volume, Sperm motility and total sperm account while the results revealed there were no significant differences in the seminal fluid parameters between infertile patients and fertile control group in semen color, pH and sperm morphology. The overall prevalence of ASA in seminal plasma among studied papulation was 16.23% (25/154). Among healthy control group anti-sperm antibody in seminal plasma were positive in 6% (3 /50) and negative in 94% (47/50) with mean concentration of anti-sperm antibody was 24.97 ± 3.7 U/ml (normal range 0 – 60 U/ml). Among infertile patients anti-sperm antibody was positive in 21.15% (22/104) whereas negative in 78.84 % (82/104). The mean concentration of anti-sperm antibody was 40.71±2.32 U/ml. The statistical analysis revealed highly significant differences (P < 0.05) in presence of ASA in seminal plasma of infertile

patients compered to normal fertile males, also concentrations of ASA in seminal plasma was significantly higher in patients compered to healthy control males. In conclusion ASA was proportionally high among patient with infertility compered to healthy fertile control group.

Introduction

It is estimated that more than million 48.5 couples that have unprotected intercourse suffer from infertility worldwide (Ombelet et al., 2008). With prevalence between 5.0%and 25.7% (Fertility, 2013). The percentage of infertile males varied from 2.5-12% globally, largest percentage of male infertility occurred in Central and Eastern Europe (8% to 12%) and Australia (8% to 9%). North America reveals rates of male infertility 4.5-6% (Ombelet et al., 2008).According the result of metaanalysis, at least 30 million men worldwide are infertile with the highest rates in Africa and Eastern Europe (Mascarenhas et al., 2012). over-all, 50% of sterility cases are due to a factor, pure woman man factor reported as 20-30% of the problem, and the rest 20-30% is caused by a combining of both female and male factors (Sharlip et al., 2002). Data from a WHO study conducted from 1994-2000 revealed that, North and West Africa had the high rates of infertility, which ranged from 4.24%-6.35%. Central and East Asia had the lowest rates of infertility, with 2.05%-3.07% of infertility cases due to male factor alone (Mascarenhas et al., 2012). A reduction in the ability of male fertility may be caused by congenital or acquired factors such as urogenital abnormalities, varicocele,

infections of the genital tract, genetic abnormalities, hormonal abnormalities, testicular failure. immunologic problems, cancer, systemic diseases, altered lifestyle, and exposure to gonadotoxic factors (Dohle et al., 2005), (Jungwirth et al., 2012). The cause of fertility disability cannot be determined in many cases, in spite of advances in diagnostic tests. the Infertility of unknown cause, it is a situation in which fertility defect occurs spontaneously or caused by an obscure or unknown cause. Infertility of unknown cause accounts about 37-58% (Moghissi & Wallach, 1983), (Jungwirth et al., 2012), (Rowe, Comhaire, Hargreave, Mellows, & Organization, 1993). The category 'unexplained male infertility' (UMI) is for infertile men reserved with infertility of unknown origin with normal semen parameters and have normal findings on clinical examination and hormonal laboratory testing and in which female infertility factors have been ruled out (Jungwirth et al., 2012).

The initial assessment of subfertile men includes medical history, physical examination, and at least two semen analyses after 12 months of unprotected intercourse (Pacey & Eiser, 2011). In approximately half of the patients, the initial assessment will identify the cause of infertility,

whereas many other patients will need to go through several complementary tests to find its cause (Wiser, Sandlow, & Köhler, 2012).

Immunologic factors are significant cause regarded as for infertility (Bohring, Krause, Habermann, & Krause, 2001). The first immunological correlation with male infertility was reported in 1954 by Wilson and Rumke with the identification of anti-sperm antibodies (Zhao, Zhao, Zhang, & Zhang, 2015), (Vazquez- Levin, et al. 2014). The prevalence of anti-sperm antibodies in infertile men varies from 9%-36% (Jiang et al., 2016) The main cause being the loss of the bloodtesticular barrier and the association with chronic inflammation (Garcia, Rubio, & Pereira, 2007). Immune infertility has been shown to be found in 15% of patients with varicocele (Rossato, Galeazzi, Ferigo, & Foresta, 2004). Anti-sperm antibodies are found in cervical mucus, seminal fluid of men, Presence of anti-sperm antibodies in seminal fluid considered risk factor for infertility (Bohring et al., 2001). Despite the fact that millions of sperm cells are present within seminal fluid, only hundreds to tens found at any one within the fallopian tube (Esteves, Schneider, & Verza Jr, 2007). Immune response to sperm able to induce male infertility; antibodies that formed against spermatozoa did not necessarily impair male fertility unless the circulating antibodies are also found within the reproductive tract and on the living spermatozoa surface.

Anti-sperm antibodies are produced in autoimmune infertility. Germ cells are normally seq uestrated from the immune system by the blood-testis barrier formed by Sertoli cells (Hjort, 1999). situations that cause damage of this barrier, like trauma, testicular surgery, varicocele, infection and inflammation as orchitis, may cause exposure of the germ cells to the immune system and production of antispermatozoa antibodies (Esteves et al., 2007), (Francavilla, Santucci, Barbonetti, & Francavilla, 2007), (Francavilla et al., 2007). The body treat the spermatozoa as foreign body. To clarify this, three hypothesizes of explanation. The first claim that spermatozoa are not display at the time of embryological development during which the immune system establishes tolerance to self-antigens (Francavilla et al., 2007). The second explanation that spermatozoa are haploid and have not the same chromosomal make-up from the somatic cells (Francavilla et al., 2007). The third hypothesis, named 'immunosuppression theory', suggests that T-suppressor lymphocytes, which suppress immune response, are activated by small amounts of spermatozoal antigens continuously leaked from the genital tract (Francavilla et al., 2007). Immediately when sperms formed during puberty, the immune system consider them as foreign cells, so they should be completely isolated from the immune system (Kayama, 2005). This isolation occurs within the testis, one of the immunologically privileged sites, by the blood-testis barrier. In other

regions of the male genital tract, the epithelial lining, probably supplemented by local a barrier. immunosuppressive is responsible for this isolation (Kayama, 2005). In spite of its immuneprivileged condition, the testis is clearly able of mounting inflammatory responses, as proved by its effective cellular and humoral defense against infections (Kipersztok et al., 2003), (DIEKMAN et al., 2000). It is well documented that the presence of antisperm antibodies can interfere with fertilization related to harmful effects embryonic development on and implantation.6,7(Verón et al., 2016), (Cui et al., 2015). It has also been Together cellular and humoral immunity have been involved in the cause of immune infertility disease (Ombelet et al., 2003). Antispermatozoa antibodies could be defined as immunoglobulins of the IgG, IgA or IgM isotype that are formed against various parts of the sperm (head, tail, mid-piece or all of them) (Yeh, Acosta, Seltman, & Immunoglobulin Doncel, 1995). subclasses IgA and IgG can be found in the ejaculates of males with antisperm autoimmunity. However, IgM looks to have no clinical effect, it is rarely detected alone or combined with IgA or IgG (Ombelet et al., 2003),(Yeh et al., 1995).

Aim of the study:

- To identify the relation between presence of antisperm antibodies and male fertility alteration in Thi-Qar governorate.

- To estimate the prevalence of the presence of antisperm antibody in Thi-Qar governorate.
- To evaluate the association between presence of ASA in idiopathic infertile male

Material and methods: The study was conducted in Thi-Qar governorate, south of Iraq. A total 154 specimens of seminal fluid were collected for study. A case-control study involve fifty (50) healthy fertile males as control group one hundred four (104) males and with fertility disorders who attending infertility center / Al-Hussein Teaching hospital, from 20/10/2016 to 16/1/2018. aged between (18-51) years with mean of age was (29.95 ± 0.68) and (50) healthy controls males aged between (23-37) years with mean age (27.58±0.51). All patient was submitted full reports about their medical status history, in addition to their infertility type (primary or secondary). Seminal fluid samples were collected by masturbation after three days of sexual avoidance. All patients and healthy control males were tested for presence of antiantibodies their spermatozoa in seminal fluid.

Ethical and Official Approval: The procedure of this research were approved and granted from the ethical research approval committee of college of medicine / university of basrah. All participants were given verbal approvals to involvement in this study.

Seminal fluid samples were collected in a private room adjacent the laboratory, in order to limit the exposure of semen to fluctuations in temperature and to control the time between collection and analysis. The were collected after a samples minimum of two days and a maximum of seven days of sexual avoidance according to WHO 2010 criteria. The man informed by clear spoken instructions about the collection of seminal fluid sample to ensure that seminal fluid sample be complete and not loss of any fraction of the sample. The following information was recorded on the report form the man's name, age and personal code number,

the period of abstinence, the date and time of collection, the completeness of difficulties the sample, any in producing the sample, and the interval between collection and the start of the analysis (WHO standards semen 2010) . Anti-Spermatozoa Antibody ELISA test used for the determination antibodies directed against human spermatozoa. This test is designed for the use with seminal plasma.

Results

1. Sorting of age in this study

In the current study the total number of males included in this study was (n=154), lo4 of them were infertile patients while the other 50 males were fertile healthy control group. with mean age (29.95±0.68) and (27.58±0.51) years respectively, also the age distributed in to three age groups and the age intervals as shown in (table 1). In the infertile patients, the highest percentage of studied patients was with in age interval 26-35 was 56 patients (53.84%). followed by age interval 15-25 was 27 patients (25.96%) and then the age interval 36-51 was 21 patients (20.19%) which represents the lowest percentage. In the fertile control group, the highest percentage within age interval 26-35 was 76% (38 patients). Followed by age interval 15-25 was 18% (9 patients) and the age group 36-51 which showed the lowest percentage 6% (3 patients) as shown in table (1). The statistical analysis revealed there was significant association (P < 0.05) among age intervals of patients and control group.

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Age groups(year)	Patients			Healthy control		
	Total	No.	%	No.	%	
15-25	36 (23.37%)	27	25.96	9	18	
26-35	94 (61.03%)	56	53.84	38	76	
36-51	24 (15.58%)	21	20.19	3	6	
Total number	154 (100%)	104	100	50	100	
Mean ± SE	lean ± SE			27.58±0.51		
\mathbf{X}^2		33.510*		63.06*		
P value		< 0.00001		< 0.00001		

2. Seminal fluid parameters and types of infertility

The presented results of seminal fluid analysis in this study showed there were significant differences (p < 0.05) between fertile control group and infertile males (patients) in seminal fluid parameters as revealed in table (2). The seminal fluid viscosity was normal in 98% of fertile healthy males in comparison to 82.69% of infertile patients with normal viscosity while 17.3% of patients have abnormal result. Liquefaction time it was normal (within 30-60 min after collection of the sample) in 100% fertile healthy males while it was 74.03% of infertile males . Semen volume was normal in 100% and 90.38% of fertile healty males and infertile males respectively. Sperm motility was normal in 100% and 75.96% of fertile healty males and infertile males respectively while it was abnormal in 24.04% of infertile patients. Sperm vitality was normal in 100% of fertile healty males while it was normal in 91.43% of infertile males. The mean of total sperm count was 259.98 ± 1.16 million sperm/ ejaculate and 180.61 ± 0.77 million sperm / ejaculate of fertile healty males and infertile males respectively. The mean of sperm concentration was 62.73 ± 0.08 million sperm/ ml for fertile healty males while it was 54.32 ± 0.13 million sperm/ ml for infertile males. The other seminal fluid parameters as semen color, seminal fluid pH and percentage of sperm normal morphology were revealed non-significant differences (P > 0.05) as shown in table (2).

Regarding the type of infertility, the primary type of infertility was the more frequent type (83.65%) compared to secondary type of infertility (16.34%), and according to

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statistical analysis the difference was significant (P < 0.05) between two types of infertility as shown in (table 2).

Table (2) Seminal fluid parameters and types of infertility in patients and control.

Semen Param eters	Fertilemales(healthycontrol)(50)	Infertile males (104)	T- tes t	X ²	P valu e	Normal WHO ranges		
Viscosi ty	(49/50) 98% within WHO normal range	(86/104) 82.69%		7. 31 6	0.00 7	Abnormal viscosity when the thread exceeds 2 cm		
liquefa ction	(50/50) 100% within (30-60)min	(77/104) 74.03%normal and (27/104) 25.96% take more than 60 min		15 .7 40	<0.0 0001	Normal within (15-60 min) when take more than 60 min. must be recorded		
Color	(50/50) 100% Gray- opalescent	(104/104) 100% Gray- opalescent		0	1	Gray- opalescent		
Semen Volum e (S.V.)	(50/50) 100% with normal semen volume Mean vol. \pm SD (4.139 \pm 0.018) ml	(94/104) 90.38% normal S.Vol. while 9.62% with S. Vol. <1.5 with mean vol. \pm SD (3.228 \pm 0.01) ml		43 .1 02	<0.0 0001	opalescent normal semen volume >1.5 ml		
рН	(50/50) 100% alkaline	(102/104) 98.07% alkaline while 1.93% (2/104) acidic		1. 01 4	o.31 4	Alkaline		
Total sperm count.	259.48 ± 1.16 million sperm	180.61 ± 0.77 million sperm	56. 35 5		<0.0 0001	$\begin{array}{c c} \text{WHO} \\ \text{(LRL)} & 39 \\ \times & 10^6 \\ \text{spermatoz} \\ \text{oa} & \text{per} \\ \text{ejaculate} \end{array}$		
Sperm count per ml	62.73 ± 0.08 million sperm	54.32 ± 0.13 million sperm	53. 46 6		<0.0 0001	WHO (LRL) 15 \times 10 ⁶ spermatoz oa per ml		
No. Males within	(50/50)100% withinnormalrange>4%withmean	(104/104) 100% within normal range >4% with mean percentage of normality (53.19 \pm 0.05%)		0	1	The lower reference		

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WHO range of normal % of sperm morph ology	percentage of normality (60.86 ± 0.06 %)					limit for normal forms is 4%		
Sperm motilit y	(50/50) 100% have PR >32% and (PR+NP) >40%	(79/104) 75.96% have PR >32% and (PR+NP) >40%		14. 34 9	<0.0 0001	PR >32% and (PR+NP) >40%		
Sperm Vitality	(50/50)100%normalvitalitypercentage >58%	91.34% (95/lo4) with normal sperms vitality >58%		4. 59 5	0.03 2	>58%		
Type of infertili ty		83.65% (87/104) with primary infertility while 16.34% (17/104) was with secondary type of infertility	92. 48 0		<0.0 0001			

3. Detection of anti-spermatozoa antibody in seminal plasma by ELISA test.

overall prevalence of ASA in seminal plasma among studied papulation was 16.23% (25/154). Among healthy control group, anti-sperm antibody in seminal plasma were positive in 6% (3 /50) and negative in 94% (47/50) with mean concentration of anti-sperm antibody was 24.97 \pm 3.7 U/ml (normal range 0 – 60 U/ml). Among infertile patients anti-sperm antibody was positive in 21.15% (22/104) whereas negative in 78.84 % (82/104). The mean concentration of anti-sperm antibody was 40.71 \pm 2.32 U/ml. The statistical analysis revealed highly significant differences (P <0.05) in presence of ASA in seminal plasma of infertile patients compered to normal fertile males, also concentrations of ASA in seminal plasma was significantly higher in patients compered to healthy control males and as shown in (3).

Table (3) presence of anti-sperm antibody in the seminal plasma in the healthy control and patients as estimated by ELISA.

ASA test result	Total number & %		le males thy control)	Infer	Infertility males		P value
		Ν	%	Ν	%		
Positive	25 (16.23%)	3	6	22	21.15		
Negative	129 (83.76%)	47	94	82	78.84		
Total	154 (100%)	50	104	104	100		
Mean		24.97	±3.7	40.71	l±2.32	3.742*	< 0.00001
conc.							
±SE							
\mathbf{X}^2		5.702	*				
P value		0.017					

4. Distribution of anti-sperm antibody in relation to age groups of infertile patients and fertile males.

The ASA was detected in 21.15% of infertile male and in 6% of normal healthy control. The majority of infertile males (50%) with positive ASA in their seminal plasma was found in age interval of 26-35 y old. followed by 27.27% in age interval 36-51 y old and then 22.72% in age interval 15-25 y old as shown in (table 4). This difference was statistically not significant (p> 0.05) among age groups.

Age distribution in fertile healthy control shows that, the highest percentage of ASA was 66.66% (2/3) detected in age interval (26-35). followed by 33.33% (1/3) in age interval (15-25). This difference was statistically not- significant (p>0.05) as shown in (table 4).

Age groups	Total	Infe	ertility			fert	ility		
	number	negativ		Positive		Negative		Positive	
		Ν	%	Ν	%	Ν	%	Ν	%
15-25	36(23.37)	22	26.82	5	22.72	8	17.02	1	33.33
26-35	94(61.03)	45	54.87	11	50	36	76.59	2	66.66
36-51	24(15.58)	15	18.29	6	27.27	3	6.38	0	0
Total	154(100)	82	53.24	22	14.28	47	30.51	3	1.94
Mean conc.±SE		32.6	59±1.84	77.2	6±2.32	19.9	91±1.94	104	.72±29.78
X2		7.75	58						
P value		0.25	56						

Table (4) distribution of anti-sperm antibody in relation to age groups infertile males and fertile males

5. Anti-spermatozoa antibody in relation to type of infertility.

Primary infertility accounted to 83.65% of patients while secondary type of infertility was (16.34%). According to the type of infertility, the majority of patients with positive ASA suffering from primary type of infertility (86.36%) but only 13.63% with secondary type of infertility (table 5). The difference was statistically significant (P<0.05). However, the mean concentration of ASA of infertile was 77.26 IU/ml compared to that among control group (32.69 IU/ml).

(Table 5.) Anti-sperm antibody in relation to types of infertility.

Infertility	Total	ASA	test result			T value	P value
type	number& %	Nega	ntive	Positive			
		Ν	%	N %			
Primary	87(83.65%)	68	82.92	19	86.36		
secondary	17(16.34%)	14	17.07	3	13.63		
Total	104(100%)	82	100	22	100		
Mean		32.69	9±1.84	77.26±2.32		10.858*	< 0.00001
conc.±SE							
\mathbf{X}^2		71.12	22*	23.273*			
P value		< 0.0	0001	< 0.000	001		

Discussion:

Patients with primary infertility represent the more frequent visitors to the fertility clinics (84%) in this study which is almost consistent with other studies in Iraq from Al-Nahrain (80.9%) (Al-Dujaily, University, Chakir, & Hantoosh, 2012) and Dohok (77.2%) (Razzak & Wais, 2002) at the north of Iraq. However, in another study from India reported a lower rate (62%) of primary infertility type (Samal, Dhadwe, Gupta, & Gupta, 2012). The findings from studies above revealed that, the primary infertility seems to be higher than secondary infertility and this will support the result found in this study.

Because the body treat the sperms as foreign body. Autoimmune infertility has long been postulated as one of the causes of subfertility (Bohring et al., 2001), (Esteves et al., 2007) and The prevalence of anti-sperm antibodies in infertile men varies from 9%-36% (Jiang et al., 2016).

Rumke and Wilson first documented that, the presence of anti-spermatozoa antibodies in infertile males, and suggest the possible potential effect of ASA in causes of infertility in males (Wilson, 1954),

In our study there was significant difference (p < 0.05) in the presence of anti-sperm antibody in seminal fluid between fertile healthy control group (6%). and infertile patients (21.15%). However, some studies reported ASA

rates lower than the figure in this study (Hossain, Islam, Aryal, & Madanes, 2007). While another study in kirkuk university almost in agreement with our finding (26.6%) of infertile patients who shows ASA in their seminal fluids plasma (p< 0.05) and considered statistically significance (Hussein, 2012). The results from this study showed that the majority of infertile patients with ASA was in age group of 25-35 years of age (50%) which is comparable to (Arora, SUDHAN, & Sharma, 1999) study which revealed that, 64% of sub-fertile patients were in the age group of 30-35. There were only 3% patients between the ages of 20 and 25 years and only 2% between the ages of 40-50 years (Arora et al., 1999)

Also this study revealed that, the positive ASA results were distributed to 86.36% with primary type of infertility and 13.63% with secondary type of infertility which is consistent with other studies reported by (Foresta et al. 2015) and in an Indian study (Khatoon, Chaudhari, Singh, & Prajapati, 2011) which included 109 infertile couples revealed that, out of which 71 were suffering from primary infertility & 38 were of secondary infertility, among those with primary infertility couples 52.12% were positive for ASA and in secondary infertility couples the incidence of ASA was 39.47% (Khatoon et al., 2011) which is almost consistent with

the finding in this study. However, another study performed by (Damianova, Dimitrova-Dikanarova, Kalaĭdzhiev, & Vatev, 1999), revealed that no significant difference in the incidence of anti-sperm antibodies among primary and secondary infertility and the highest incidence of anti-sperm antibodies amongst patients with primary unexplained infertility (Damianova et al., 1999) that in agreement with this study where there was non-significant difference in the percentage of ASA in primary and secondary infertility.

References

Al-Dujaily, S. S., Chakir, W. K., & Hantoosh, S. F. (2012). Direct antisperm antibody examination of infertile men. *Global Journal of Medical Research*, *12*(3).

Arora, P., SUDHAN, M. D., & Sharma, R. K. (1999). Incidence of anti-Sperm antibodies in infertile male population. *Medical Journal Armed Forces India*, *55*(3), 206–208.

Bohring, C., Krause, E., Habermann, B., & Krause, W. (2001). Isolation and identification of sperm membrane antigens recognized by antisperm antibodies, and their possible role in immunological infertility disease. *Molecular Human Reproduction*, *7*(2), 113–118.

Cui, D., Han, G., Shang, Y., Liu, C., Xia, L., Li, L., & Yi, S. (2015). Antisperm antibodies in infertile men and their effect on semen parameters: a systematic review and meta-analysis. *Clinica Chimica Acta*, 444, 29–36.

Damianova, V., Dimitrova-Dikanarova, D., Kalaĭdzhiev, S., & Vatev, I. (1999). The incidence of sperm antibodies in patients included in a program of assisted reproduction. *Akusherstvo i Ginekologiia*, *38*(2), 31–33.

DIEKMAN, A. B., NORTON, E. J., WESTBROOK, V. A., KLOTZ, K. L., NAABY- HANSEN, S., & HERR, J. C. (2000). Anti- sperm antibodies from infertile patients and their cognate sperm antigens: a review. Identity between SAGA- 1, the H6- 3C4 antigen, and CD52. *American Journal of Reproductive Immunology*, *43*(3), 134–143.

Dohle, G. R., Colpi, G. M., Hargreave, T. B., Papp, G. K., Jungwirth, A., Weidner, W., & Infertility, E. A. U. W. G. on M. (2005). EAU guidelines on male infertility. *European Urology*, *48*(5), 703–711.

Esteves, S. C., Schneider, D. T., & Verza Jr, S. (2007). Influence of antisperm antibodies in the semen on intracytoplasmic sperm injection outcome. *International Braz j Urol*, *33*(6), 795–802.

Thi-Qar Medical Journal (TQMJ):Vol.(18),No.(2),2019Web Site: https://jmed.utq.edu.iqISSN (Print):1992-92 18, ISSN (online):1992- 92 18DOI: https://doi.org/10.32792/utq/utjmed/18/2/8

Fertility, N. (2013). Assessment and treatment for people with fertility problems. *London: National Institute for Health and Care Excellence*.

Francavilla, F., Santucci, R., Barbonetti, A., & Francavilla, S. (2007). Naturallyoccurring antisperm antibodies in men: interference with fertility and clinical implications. An update. *Front Biosci*, *12*, 2890–2911.

Garcia, P. C., Rubio, E. M., & Pereira, O. C. M. (2007). Antisperm antibodies in infertile men and their correlation with seminal parameters. *Reproductive Medicine and Biology*, *6*(1), 33–38.

Hjort, T. (1999). Antisperm antibodies. Antisperm antibodies and infertility: an unsolvable question? *Human Reproduction (Oxford, England)*, *14*(10), 2423–2426. https://doi.org/10.1093/humrep/14.10.2423

Hossain, A., Islam, N., Aryal, S., & Madanes, A. (2007). The prevalence of circulating anti sperm antibody (ASA) in infertile population representing of all etiologies. *Middle East Fertility Society Journal*, *12*(1), 27.

Hussein, M. A. A. R. H. N. (2012). Prevalence of Antisperm Antibodies in Asthenospermic Infertile Male. *Kirkuk University Journal for Scientific Studies*, 7(1), 1–7.

Jiang, Y., Cui, D., Du, Y., Lu, J., Yang, L., Li, J., ... Bai, X. (2016). Association of anti-sperm antibodies with chronic prostatitis: a systematic review and metaanalysis. *Journal of Reproductive Immunology*, *118*, 85–91.

Jungwirth, A., Giwercman, A., Tournaye, H., Diemer, T., Kopa, Z., Dohle, G., ... Infertility, E. A. U. W. G. on M. (2012). European Association of Urology guidelines on Male Infertility: the 2012 update. *European Urology*, 62(2), 324– 332.

Kayama, K. (2005). Immunologic tests: anti-sperm antibody a review. *Japanese J Clinical Medicine*, *63*(7), 581–584.

Khatoon, M., Chaudhari, A. R., Singh, R., & Prajapati, S. (2011). Antisperm antibodies in primary and secondary infertile couples of Central India.

Kipersztok, S., Kim, B. D., Morris, L., Drury, K. C., Williams, R. S., & Rhoton-Vlasak, A. (2003). Validity of a rapid assay for antisperm antibodies in semen. *Fertility and Sterility*, 79(3), 522–528.

Mascarenhas, M. N., Flaxman, S. R., Boerma, T., Vanderpoel, S., & Stevens, G. A. (2012). National, regional, and global trends in infertility prevalence since 1990: a systematic analysis of 277 health surveys. *PLoS Medicine*, *9*(12), e1001356.

Moghissi, K. S., & Wallach, E. E. (1983). Unexplained infertility. *Fertility and Sterility*, *39*(1), 5–21.

Thi-Qar Medical Journal (TQMJ):Vol.(18),No.(2),2019Web Site: https://jmed.utq.edu.iqISSN (Print):1992-92 18, ISSN (online):1992- 92 18DOI: https://doi.org/10.32792/utq/utjmed/18/2/8

Ombelet, W., Cooke, I., Dyer, S., Serour, G., & Devroey, P. (2008). Infertility and the provision of infertility medical services in developing countries. *Human Reproduction Update*, *14*(6), 605–621.

Ombelet, W., Deblaere, K., Bosmans, E., Cox, A., Jacobs, P., Janssen, M., & Nijs, M. (2003). Semen quality and intrauterine insemination. *Reproductive BioMedicine Online*, *7*(4), 485–492.

Pacey, A. A., & Eiser, C. (2011). Banking sperm is only the first of many decisions for men: What healthcare professionals and men need to know. *Human Fertility*, *14*(4), 208–217.

Razzak, A. H., & Wais, S. A. (2002). *The infertile couple: a cohort study in Duhok, Iraq.*

Rossato, M., Galeazzi, C., Ferigo, M., & Foresta, C. (2004). Antisperm antibodies modify plasma membrane functional integrity and inhibit osmosensitive calcium influx in human sperm. *Human Reproduction*, *19*(8), 1816–1820.

Rowe, P. J., Comhaire, F. H., Hargreave, T. B., Mellows, H. J., & Organization, W. H. (1993). *WHO manual for the standardized investigation and diagnosis of the infertile couple*.

Samal, S., Dhadwe, K., Gupta, U., & Gupta, N. K. (2012). Epidemiological study of male infertility. *Indian Medical Gazette*, *5*, 174–180.

Sharlip, I. D., Jarow, J. P., Belker, A. M., Lipshultz, L. I., Sigman, M., Thomas, A. J., ... Damewood, M. D. (2002). Best practice policies for male infertility. *Fertility and Sterility*, 77(5), 873–882.

Vazquez- Levin, M. H., Marín- Briggiler, C. I., & Veaute, C. (2014). Antisperm antibodies: invaluable tools toward the identification of sperm proteins involved in fertilization. *American Journal of Reproductive Immunology*, 72(2), 206–218.

Verón, G. L., Molina, R. I., Tissera, A. D., Estofan, G. M., Marín-Briggiler, C. I., & Vazquez- Levin, M. H. (2016). Incidence of sperm surface autoantibodies and relationship with routine semen parameters and sperm kinematics. *American Journal of Reproductive Immunology*, *76*(1), 59–69.

Wilson, L. (1954). Sperm agglutinins in human semen and blood. *Proceedings of the Society for Experimental Biology and Medicine*, 85(4), 652–655.

Wiser, H. J., Sandlow, J., & Köhler, T. S. (2012). *Causes of male infertility In: Parekattil JS, Agarwal A.(eds). Male Infertility: Contemporary Clinical Approaches, Andrology, ART & Antioxidants.* New York: Springer New York.

Yeh, W.-R., Acosta, A. A., Seltman, H. J., & Doncel, G. (1995). Impact of immunoglobulin isotype and sperm surface location of antisperm antibodies on

Thi-Qar Medical Journal (TQMJ):Vol.(18),No.(2),2019 Web Site: https://jmed.utq.edu.iq Email:utjmed@utq.edu.iq ISSN (Print):1992-92 18, ISSN (online):1992-92 18 DOI: https://doi.org/10.32792/utg/utjmed/18/2/8 fertilization in vitro in the human. Fertility and Sterility, 63(6), 1287–1292.

Zhao, Y., Zhao, E., Zhang, C., & Zhang, H. (2015). Study of the changes of acrosomal enzyme, nitric oxide synthase, and superoxide dismutase of infertile patients with positive antisperm antibody in seminal plasma. Cell Biochemistry and Biophysics, 73(3), 639-642.

التقييم السير ولوجي لوجود اضداد النطف في الرجال العقيمين في محافظة ذی قار عباس فاضل غضبان السعيدي (دكتوراه، ماجستير) فرع الأحياء المجهرية \ كلية الطب- جامعة البصرة حسن جابر حسونی (دكتوراه ، ماجستير) فرع الاحياء المجهرية) كلية الطب- جامعة البصرة الدكتورة أيناس صالح الخياط استاذ مساعد \ فرع النسائية - كلية الطب جامعة ذي قار

الخلاصة: أجريت هذه الدراسة (case control study) في محافظة ذي قار، جنوب العراق، شملت ما مجموعه ١٠٤ شخصاً (males) (منهم ١٠٤ مرضى يعانون من العقم، في حين كان مجموعة السيطرة تتكون من ٥٠ من الرجال الخصبين)، كان المرضى من الرجال الذين يراجعون مركز العقم في مستشفى الامام الحسين التعليمي من ٢٠١٦/١٠/٢٠ إلى ٢٠١٨/١/١٦ ،والذين تتراوح أعمار هم بين (١٨- ٥١) سنة مع متوسط العمر (٢٩,٩٥ ± ٢٩,٩٠) واما بالنسبة الى (٥٠) رجل الاخرين الذين يمثلون مجموعة السيطرة فكان معدل أعمارهم تتراوح بين (٢٣-٣٧) سنة مع متوسط العمر كان (٢٧,٥٨ ± ٢٥,٥١). قدّم جميع المرضى تقارير كاملة عن تاريخ حالتهم الطبية، بالإضافة إلى نوع العقم (الابتدائي أو الثانوي). تم جمع عينات السائل المنوي بعد ثلاثة أيام من الامتناع ألجنسي. تم فحص جميع المرضى ومجموعة السيطرة بطريقة (seminal fluid analysis) و حسب ضوابط كتاب منظمة الصحة العالمية في اجراء فحص السائل المنوى لسنة ٢٠١٠ الطبعة الخامسة. بالإضافة الى ذلك تم فحص كلتا المجمو عتين بطريقة ELISA للتحرى عن وجود الأجسام المضادة للنطف (ASA) في السائل المنوي

. أظهر نتائج الفحص العام للسائل المنوي أن هناك اختلافات كبيرة (P < ٠,٠٠) بين مجموعة السيطرة والمرضى في (لزُوجة السائل المنوي، وقت التسييل، حجم السائل المنوى، حركية الحيوانات المنوية وعدد الحيوانات المنوية الكلي في حين

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كشفت النتائج أنه لم تكن هناك اختلافات ذات قيمة إحصائية في صفات السائل المنوية (لون السائل المنوي، ودرجة الحموضة (pH) وشكل الحيوانات المنوية.

في حين أظهرت نتائج الفحص السيورولوجي بطريقة ELISA للتحري عن الحيوانات المنوية ما يلي

ان الانتشار العام ASA في البلازما المنوية في ١٥٤ رجل (كلتا المجموعتين التي دخل الدراسة من الرجال) كانت ١٦,٢٣٪ (١٥٤/٢٥). وقد كانت نتيجة فحص وجود الجسام المضادة للحيوانات المنوية في بلازما السائل المنوي في مجموعة السيطرة إيجابية في ٢٦, (١٠٤/ (١٥٤/ (١٠٤)) مع متوسط تركيز (السيطرة إيجابية في ٢٢, (٣ /٥٠) وسلبية في ٩٤٪ (٧٠/ ٢٠) مع متوسط تركيز (السيطرة إيجابية في ٢٦, (٣ /٥٠) وسلبية في ٢٤٪ (٢٠٤ / ٢٤) مع متوسط تركيز (١٠٤/ ٢٤) لما بالنسبة السيطرة إيجابية في ٢٠, (٣ /٥٠) وسلبية في ٢٤٪ (٢٠٤ / ٢٤) مع متوسط تركيز (١٠٤ / ٢٢) مع متوسط تركيز (١٠٤ / ٢٢) مع متوسط تركيز الأجسام الى المرضى الذين يعانون من العقم فقد كان فحص ASA إيجابي في ١٠٤٪ (٢٠٤ / ٢٠٤). وكان متوسط تركيز الأجسام الى المرضى الذين يعانون من العقم بالمقارنة إلى الذكور الخصبين، كما كانت المنوية من المرضى الذين يعانون من العقم بالمقارنة إلى الذكور الخصبين، كما كانت من المرضى من المرضى الذين يعانون من العقم بالمقارنة إلى الذكور الخصبين، كما كانت اختلافات كبيرة ASA في البلازما المنوية ما من المرضى الذين يعانون من العقم بالمقارنة إلى الذكور الخصبين من المنوية من المرضى الذين يعانون من العقم بالمقارنة إلى الذكور الخصبين، كما كانت من المرضى الذين يعانون من العقم بالمقارنة إلى الذكور الخصبين، كما كانت مركيز من المرضى الذين يعانون من العقم بالمقارنة إلى مرضي. الملخص ان تركيز من محموعة السيطرة المرضى الذين يعانون من المرضى الذين يعانون من العقم بالمقارنة إلى مجموعة السيطرة إلى محموعة السيطرة إلى المورية الم ما مرضي الذين يعانون من العقم بالمقارنة إلى محموعة السيطرة إلى محموعة السيطرة المرضي ما المرضي الذين يعانون من المورين ما المنوية أعلى محموين المن المورية ألى محموية ألى محموية ألى محموية ألى المرضي المرضي المورية إلى محموية السيطرة إلى محموية السيطرة إلى المرضي الذين يا مرك مى محموية السيطرة إلى محموية السيطرة إلى الموري الم ما مرك مى الذين محموية السيطرة مي الموري ما الموري ما الموري ما الموري مى الموري مى الموري مى الموري ما الموري ما الموري ما الموري مى المو