By *Plasmodium berghei* Khalid Majeed Dakhel Assistance prof.*

SUMMARY

Twenty mice were injected i.p. with $1 \times 10^5 P$. *berghei* infected red cells . Blood smears of these mice were prepared daily till the mice were died . Mice were categorized in three groups depending on similar type of course of parasitaemia after inoculation . Seven mice of Group A (A₁ – A₇) showed similar type of behavior and died due to low parasite infection . Another group of seven mice (B₁ to B₇) showed a gradual increase in parasitaemia till mice died due to high infection . Group C (C₁ to C₆) smears showed a vary rise and fall in infection and all the mice were cleared of parasite infection .

The antibody titre as detected by IFA, observed in these groups was 1:2048, 1:1024 and 1:4096 in groups A, B and C respectively. The group C which showed protective immunity against *P*. *berghei* infection depicted strong fluorescence against normal erythrocyte membrane too. The fluorescence against normal RBC_S membrane was observed very weak in group A, whereas it was negligible in group B.

INTRODUCTION

Malaria continues to cause morbidity and mortality on a large scale in tropical countries . According to the report given by WHO [1] , human malaria parasite causes 300 - 500 million clinical cases which include young children under 5-6 years of age , pregnant women and non – immune travelers . Annual mortality caused by malaria ranges from 0.5 - 2.5million including over a million death in children aged under five [2].

Earlier studies have greatly influenced the knowledge about invasion of parasite in red cells [3] and the pioneer work suggested the role of receptors in recognition of susceptible host cells by merozoites for invasion .[4]

In vitro studies of P . *knowlesi* have shown the behavior of merozoites to be even more complex, since the process of invasion involves various sequential changes in shapes of both host and parasite cells [5]

A study was described that the interactions between *P* . *falciparum* merozoites and human RBC is mediated

by specific parasite proteins and sialoglycoproteins (SGPS) on the surface of the host cell.[6]

The first *plasmodium* molecule found to be located in the microneme organelles of the merozoite is the 135 kDa P. Knowlesi Duffy binding proteins (pk DBP) [7] . The gene encoding this proteins was subsequently used as a probe to clone the corresponding gene from P.vivax (Pv DBP) [8]. It was that the invasion suggested of and glycophorin B deficient Α erythrocytes by P. falciparum as trypsin or neuraminidase treated glycophorin B deficient cells were refractory to the parasite EBA-175, a parasite protein appears to bind to glycophorin A . [9,10] suggested the EBA as transmembrane proteins characterized by conserved cysteine rich domain expressed in the micronemes of invasive merozoites . Strong evidences have implicated EBA – 175 as being a parasite – ligand used in the recognition of human RBC by p. falciparum merozoite [11]. Merozoite surface protein 2 (MSP-2) a highly polymeric in nature, generates

^{*} Community Health Dept. Nassiriah Technical Institute

antiparasite humoral response in mice [12]. Vaccination of animals against leads to formation sporozoute of antibodies that inhibit sporozoute motility and block the invasion of blood vessels by sporozoute [13]. A study found that plasmodium - infected ervthrocvte release a low molecular weight soluble factor that inhibits the activation of macrophages and dedritic cells [14]. The present work is an attempt to understand the preferential invasion of reticulocytes by the parasite in p. berghei infection.

MATERIAL & METHODS

Parasite Normal mice were injected intraperitoneally (i.p.) with parasitized red cell in citrate saline and the course of parasitaemia was monitored .

Accordingly the mice were grouped in different categories . NK 65 , a lethal strain of *P* . *berghei* was maintained in Balb / C mice swiss white mice . 20 mice each of 20-25 gms . were inoculated with $1x10^5$ parasitized cells and daily course of infection was monitored by Giemsa stained blood smears . The percentage of total number of red blood cells :

Collection of Blood

The serum was collected in two ways :

- 1- A few drops of blood were collected from each mouse separately. It was allowed to clot at room temperature for one hour. The clot was broken and serum was isolated.
- 2- Mice were anesthetized by diethyl ether and bled by jugular vein incision. The blood from broken clot was then centrifuged at 1,000 g for 15 minutes and the serum was aspirated and collected.

Statistics analysis :

Chi (X^2) test was used for statically analysis in present study.

Indirect Fluorescent Antibody (IFA) test IFA test was done according to slightly modified method (1972) [13]. The slides were counter stained with Evan's blue and studied under UV. Fluorescence microscope (Leica DMILS, Germany).

RESULTS

Group A :

The group contained seven mice $(A_1 to$ A₇). All the mice of this group died due to infection and maximum parasitaemia remained between (24% - 60%)in most of mice (Fig. 1). One of mice had only 7% maximum parasitaemia . The parasitaemia ranged between (3 - 15%) till day 10 and both reticulocytes and erythrocytes were infected . One mice died on day 12, other two on day 14, and one on day 16. In the remaining three mice the parasitaemia further increased and reached about 48% to 60% followed bv increase in reticulocytosis and decrease in parasitaemia . These mice were found dead due to infection on day 24, 26 and 28.

Group B :

This group also contained seven mice (B₁ to B₇) but on the contrary to group A the animals died to high parasitaemia (Fig . 2) . Maximum parasitaemia ranged between 58% to 82%. Both erythrocytes and reticulocytes were infected but with gradual increase in parasitaemia and appearance of reticulocytes . In two of the mice of this group more than 80% cell were infected. The animals died because of high parasitaemia between day 13 to day 17. Group C:

This group contained 6 mice $(C_1 \text{ to } C_6)$. In this group the parasite was observed on 14th day of post inoculation in 4 mice . Whereas, two mice became infected on 5th day. There was Zig – Zag course of parasitaemia . In all the mice initially most of the erythrocytes were infected along with a few reticulocytes (Fig. 3) with the gradual increase in parasitaemia , reticulocytosis occurred and the post migrated from red cells to the reticulocytes and finally the parasite was exclusively confined to the reticulocytes . An increase in number of WBC_s were observed during later days. Futher smears showed decrease in parasitaemia between day 15 and day 20 and finally no parasite was observed in

the smear of this group after 25th day . Indirect Fluorescent Antibody (IFA) test: IFA test was employed to detect the presence of antibodies in parasitized mice . The IFA slides were counter stained with Evan's blue to avoid any non-specific reaction . The appearance of antimalarial antibodies in mice of different groups and the titre of antibodies are depicted in Table 1 . Group A :

Antibodies appeared in one of the mice on day 9 whereas , on 10^{th} day all the mice showed positive titre of 1 : 32 (Fig. 4) . Sera collected on day prior to the death of infected animals showed varying IFA titre . Four mice (A₁, A₂, A₆ and A₇) showed varying IFA titre between 1 : 256 and 1 : 1024 while others showed 1 : 2048 (Fig. 5).

IFA that was significant (P < 0.05) with the infection percentage .

Group B :

In this group antimalarial antibodies were observed on day 9^{th} in 2 mice (B₁ to B₄) whereas, by next day appeared in other 5 mice (Fig. 6). The IFA titre ranged between 1 : 64 which died on day 13^{th} and 1 : 512 which died on day 15^{th} an 1 : 1024 which died on 16^{th} and 17^{th} day (Fig. 7).

NO or almost negligible fluorescence was observed with normal red cell antigen . no significant (P < 0.05) between IFA and infection percentage . Group C :

The antimalarial antibody was observed on day 10th post inoculation in all the 6 mice (Fig. 8). IFA titre of 1: 4096 was observed in the sera of these mice when sacrificed on 36th day . The IFA titre of this group showed in (Fig. 9). A very strong IFA reaction was observed against all the stages of the parasite in case of parasitized erythrocytes or reticulocytes. The reaction with the host cell antigen revealed that more fluorescence observed with was erythrocytes as compared with the reticulocytes.

IFA that was significant (P < 0.05) with the infection percent .

DISCUSSION

Susceptibility of Balb / C strain of white swiss mice to NK – 65 strain of *P.berghei* as an asynchronous parasite it was well established the suscep [14].

Preferential invasion of reticulocytes by merozoites of P . berghei was also observed by others [15]; [16] ;[17] . P. vivax invades reticulocytes exclusively and reticulocyte binding gene expressed PvRBP-1 and PvRBP-2, two proteins which are shown to be responsible for preferential invasion of P. vivax in reticulocytes . P . berghei has shown preference for reticulocytes , however , PvRBP gene probes did not cross hybridise to DNA of P . falciparum , P . knowlesi and P . berghei [18] .

In Vivo reticulocytes mature to red cells within 24-36 hours after release from bone marrow into circulation [19]. During the maturation process of ervthrocytes a number of constituent proteins from the membranes of the reticulocytes are lost [20]; [21]; [22]. The phenomenon of invasion of red blood cells by the parasite is a multistep process involving attachment of the host cell and internalization of the parasite [23]; [24]. The relationship between age of the host cell and invasion bv P.falciparum indicated that voung erythrocytes in particular have an increased susceptibility to the parasite [25] . In P . berghei the parasite reticulocytes parasitizes preferably initially and exclusively later, if it remains in the host cell for 2-3 weeks or more.

Although the IFA is useful in estimating exposure to malaria, previous work has indicated that many antibodies directed against some parasite antigens detected by IFA, depicted appearance of antibodies between 7th to 10th day of post inoculation. There was a gradual increase in antibody level in different mice. Some mice died due to low parasitaemia with IFA titre of 1 : 2048 after 24 days whereas IFA titre of 1 : 1024 was observed in some mice which died due to high parasitaemia between

16th and 17th day , or some mice recovered from infection and the parasite was not observed in the smear with IFA titre of 1 : 4096 . An increase in WBC'S was observed during later days depicting some role of cell mediated immunity along with humoral

immunity, both playing a major role in protection.

Finally humoral and cell mediated immunity along with some genetic factors may be responsible for protective immunity in these mice.

Table1 : Appearance of antimalarial antibodies in mice of different groups and the tutre of antibody .

Group	Antibody appearance		Maximum Antibody Titus in
	Day	No . of mice	Antibody Titre in sera collected *
Α	9 10	1 6	1:2048
В	7 8	2 5	1:1024
С	10	6	1 : 4096



Fig. 1 : Course of P. berghel in mice resulting in death of animals due to low parasitaemia.

Thi-Qar Medical Journal (TQMJ): Vol(3) No(1):2009(57-66)



Fig. 2 : Course of P. berghei in mice resulting in death animals due to high parasitaemia



Fig. 3 : Course of P. berghei infection in mice and subsequent protection of the animals



Fig. 4 : Histogram showing antimalarial antibodies in the sera of mice of group A. Animals showing 1 : 32 or more titre were positive.



Fig. 5 : Histogram showing IFA titre in the sera samples collected in group A mice.





Fig. 6 : Histogram showing antimalarial antibodies in the setup of mice of group B. Animals showing 1 : 32 or more titre were positive.



Fig. 7 : Histogram showing IFA titre in the last sera samples collected in group B mice.



Fig. 8 : Histogram showing antimalarial antibodies in the sera of mice of group C. Animals showing 1 : 32 or more titre were positive.



Fig. 9 : Histogram showing IFA titre in sera samples of group C mice.

REFERENCES

- 1- World Health organisation. Rolling back malamina . WHO World Health Report (1999) , 49-64.
- 2- White, N.J., Nosten, F., Looareesuwan, S., Watkins, W.M., Marsh, K., Snow, R.W., Kokwaro, G., Ouma, J., Hein, T.T., Moltneux, M.E., Taylor, T.E., Newbold, C.I., Buebush, T.K., Danis, M., Greenwood, B.M., Anderson, R.M. and Ollirao, P. (1999). Averting a malaria disaster. *Lancet*, 353, 1965-1967.
- 3- Ball, E.G., Anfinsen, C.B., Geiman, Q.M., Mckee, R.W. and Ormsbee, R.A. (1945). In vitro growth and multiplicaton of the malarial parasite, *P.knowlesi.Science*, 101, 542 544.
- 4- McGhee, R.B. (1951). The adaptation of the avian malaria parasite *P.lophurae* to a continuous existence in baby mouse. *J.Infect.Dis.*, 88, 86 97.
- 5- Dvorak , J.A., Miller , L.H ., Whitehouse , H.C.and Shiroishi , T.(1975). Invasion of erythrocytes by malaria merozoites. *Science* , 187 , 748 750.
- 6- Rodriguez, M.H.and Jungery, M.(1987).Glycoprotein recognition meiates attachment of *P.chabaui* to mouse erythrocytes.*Infect.Immun*., 55, 187 192
- 7- Adams , J.H ., Hudson , D.E ., Torii , M ., Ward , G.E ., Wellens , T.E ., Aikawa , M.and Miller , L.H.(1990). The duffy receptor family of *plasmodium knowesi* in located within the micronemes of invasion merozoites. *Cell* , 63 , 141 153.
- 8- Fang , X ., Kaslow , D.C ., Adams , J.H.and Miller , L.H.(1991). Cloning of the *plasmodium vivax* Duffy receptor. *Mol.Biochem.Parasitol* ., 44 , 125 132.
- 9- Dolan, S.A., Proctor, J.L., Alling, D.W., Okubo, Y., Wellems, T.E.and Miller, L.H.(1994).Glycophorin B as an EBA 175 indepenent *plasmodium falciparum* receptor of human erythrocytes.*Mol.Biochem.Parasitol.*, 64, 55 63.
- 10- Kappe , S.H ., Curley , G.P ., Neo , A.R ., Dalton , J.P.and Adams , J.H.(1997). Erythrocyte binging protein homologues of rodent malaria parasites. *Mol.Biochem.Parasital* ., 89 , 137 148.
- 11- Rodriguez, L.E., Urquiza, M., Ocampo, M., Suarez, J., Curtidor, H., Guzman, F., Vargas, L.E., Trivinos, M., Rosas, M.and Patarroyo, M.E.(2000).*Plasmodium falciparum* EBA 175 KDa protein peptides which bind to human red blood cells.*Parasitology*, 120, 255 235.
- 12- Lawrence , N ., Stowers , A ., Mann , V ., Taylor , D.and Saul , A.(2000).Recombinant chimeric proteins generated from conserved regions of *Plasmodium falciparum* merozoite surface proteins 2 generate antiparasite humoral responses in mice.*Parasite immunology*, 22, 211 221.
- 13- Vanderberg JP , Frevert U, intrarital microscopy demonstrating antibody mediated immobilistation of P. berghen sporozoute int . J . parasitol . 2004 . Aug ; 34 (9) : 991-6.
- 14- J.C. Hafalla , U.Rai U , A. morrot , D. Bernal Rublo , F.zavala and A. Rudriquez . priming of CD_8T T cell responses following immunization with heat killed plasmodium spurozutes . European journal of immunotogy .(2006) . may ; 36 (5) :179-86.
- 15- Ramakrishnan ,S.P. and Prakash , S.(1950).Studies on *P.berghei* n.sp.Vinke and Lips (1948).Morphology , periodicity an pathogenicity in blood induce infection in mice , rats and garden squirrels.*Indian J.Malariol* ., 4 , 369 375.
- 16- Janse, C.J., Mons, B., Croon, J.J.A.B.and Vander Kaay, H.J.(1984).Long term in vitro culture of P.berghei and preliminary observations on gametocytogenesis.*Int.J.parasitol.*, 14, 317 – 320.
- 17- Janse , C.J ., Boorrsma , E.G ., Ramesar , J ., Grobbee , M.J.and Mons , B.(1989).Host cell specificity and schizogony of *P.berghei* under different in vitro conditions.*Int.J.parasitol* ., 19 , 509 514.

- 18- Galinski , M.R ., Medina , C.C ., Ingravallo , P ., Barnwell , J.W.(1992). reticulocyte – binding protein complex of *P.vivax* merozoites.Cell 69 , 1213 – 1226.
- 19- Gronowicz, G., Swift, H.and Steck, T.L.(1984). Maturation of the reticulocyte in vitro. *J. cell.sci.*, 71, 177 197.
- 20- Patel , V.P ., Ciechanover , A ., Platt , O.and Lodish , H.F.(1985).Mammilian reticulocytes lose adhesion to fibronection during maturation to reticulocytes.*Proc.Natl.Acad.Sci* ., USA , 82 , 440 444.
- 21- Rapoport, S.M.(1986). The reticulocyte. Boca Raton Florida : CRC Press.
- 22- Lazarides , E.(1987). From genes to structural morphogenesis : the genesis and epigenesis of a red blood cell. *Cell* , 51 , 345 356.
- 23- Ward , G.E ., Chitnis , C.E.and Miller , L.H.(1994). Strategies for intracellular survival of microbes (ed.Russel.D .) Saunders , 155 190.
- 24- Wilson, R.S.M.(1990).Biochemistry of red cell invasion.Blood cells, 16, 237 252.
- 25- Pasvol, G., Weatherall, D.J.and Wilson, R.J.M.(1979). The increased susceptibility of young red cells to invasion by the malarial parasite. *British Journal of Haematology*, 45, 285 295.

تأثير الاجسام المضادة على كريات الدم الحمراء المهاجمة بواسطة الملاريا P. berghei

د.خالد مجید داخل استاذ مساعد *

المستخلص

حقن ٢٠ حيوانا مختبريا بواسطة 10x1 كريات دم حمراء مصابة ب P . berghei و اخذت مسحات دم لهذه الحيوانات يوميا ولحين هلاك هذه الحيوانات وقسمت هذه الحيوانات الى ثلاثة مجاميع اعتمادا على الاستجابة للإصابة بعد الحقن وهي مجموعة A $(A_7 - A_1)$ اعطت نفس السلوك وقد تم هلاك هذه الحيوانات الاستجابة للإصابة بالطفيليات القليلة . المجموعة B $(A_7 - A_1)$ اعطت نفس السلوك وقد تم هلاك هذه الحيوانات نتيجة للإصابة بالطفيليات القليلة . المجموعة B $(A_7 - A_1)$ اعطت نفس السلوك وقد تم هلاك هذه الحيوانات ولاستجابة للإصابة مع الحقن وهي مجموعة B $(A_7 - A_1)$ اعطت نفس السلوك وقد تم هلاك هذه الحيوانات ولاستجابة للإصابة بالطفيليات القليلة . المجموعة B $(A_7 - A_1)$ اعطت زيادة تدريجية في نسبة الاصابة بالطفيليات نتيجة للإصابة مالطفيليات القليلة . المجموعة B ولي المحتر زيادة كبيرة وإصابة كاملة وكانت جميع الحيوانات ولحين هلاك الحيوانات . مجموعة C (C₆ - C₁) اعطت زيادة كبيرة وإصابة كاملة وكانت جميع الحيوانات ولحين ملاك الحيوانات . مجموعة C (C₆ - C₁) اعطت زيادة كبيرة وإصابة مالم وكانت جميع الحيوانات .

نسبة الاجسام المضادة حددت بواسطة اختبار فحص الاجسام المناعية (IFA) وقد لوحظت النتائج التالية : C , B , A وعلى التوالي . وقد اعطت المجموعة C , B , A وعلى التوالي . وقد اعطت المجموعة C , B ، مناعة كمناعة كبيرة جداً للإصابة بالطفيليات وقد كانت A ضعيفة جداً .

^{*} قسم صحة مجتمع – المعهد التقنى / ناصرية